

# Ecology of the Freshwater Pearl Mussel

*Margaritifera margaritifera*



Conserving Natura 2000 Rivers  
Ecology Series No. 2



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# Conserving Natura 2000 Rivers

This account of the ecology of the freshwater pearl mussel (*Magaritifera margaritifera*) has been produced as part of **Life in UK Rivers** – a project to develop methods for conserving the wildlife and habitats of rivers within the Natura 2000 network of protected European sites. The project's focus has been the conservation of rivers identified as Special Areas of Conservation (SACs) and of relevant habitats and species listed in annexes I and II of the European Union Directive on the Conservation of Natural Habitats and of Wild Fauna and Flora (92/43/EEC) (the Habitats Directive).

One of the main products is a set of reports collating the best available information on the ecological requirements of each species and habitat, while a complementary series contains advice on monitoring and assessment techniques. Each report has been compiled by ecologists who are studying these species and habitats in the UK, and has been subject to peer review, including scrutiny by a Technical Advisory Group established by the project partners. In the case of the monitoring techniques, further refinement has been accomplished by field-testing and by workshops involving experts and conservation practitioners.

**Life in UK Rivers** is very much a demonstration project, and although the reports have no official status in the implementation of the directive, they are intended as a helpful source of information for organisations trying to set 'conservation objectives' and to monitor for 'favourable conservation status' for these habitats and species. They can also be used to help assess plans and projects affecting Natura 2000 sites, as required by Article 6.3 of the directive.

As part of the project, conservation strategies have been produced for seven different SAC rivers in the UK. In these, you can see how the statutory conservation and environment agencies have developed objectives for the conservation of the habitats and species, and drawn up action plans with their local partners for achieving favourable conservation status.

Understanding the ecological requirements of river plants and animals is a prerequisite for setting conservation objectives, and for generating conservation strategies for SAC rivers under Article 6.1 of the Habitats Directive. Thus, the questions these ecology reports try to answer include:

- What water quality does the species need to survive and reproduce successfully?
- Are there other physical conditions, such as substrate or flow, that favour these species or cause them to decline?
- What is the extent of interdependence with other species for food or breeding success?

For each of the 13 riverine species and for the *Ranunculus* habitat, the project has also published tables setting out what can be considered as 'favourable condition' for attributes such as water quality and nutrient levels, flow conditions, river channel and riparian habitat, substrate, access for migratory fish, and level of disturbance. 'Favourable condition' is taken to be the status required of Annex I habitats and Annex II species on each Natura 2000 site to contribute adequately to 'favourable conservation status' across their natural range.

Titles in the Conserving Natura 2000 Rivers ecology and monitoring series are listed inside the back cover of this report, and copies of these, together with other project publications, are available on the project website: [www.riverlife.org.uk](http://www.riverlife.org.uk).



## Introduction

The freshwater pearl mussel, *Margaritifera margaritifera*, has been exploited in Britain since pre-Roman times. Indeed, its occurrence here may have been an important factor in inducing the first Roman invasion of Britain in 55BC (Cosgrove *et al.* 2000a). Scottish pearls were traded in Europe during the 12th Century, and commercial exploitation on a much larger scale developed during the 16th Century across Britain and Ireland, when river bailiffs were employed to ensure that all valuable pearls were kept for the King.

Many people worked in the industry during the 19th Century, but this level of exploitation was apparently unsustainable and the fishery declined sharply. There was a constant but small-scale pearl fishery, traditionally plied by travelling people, until the mussel was afforded complete legal protection in 1998. The pearl mussel is now seriously endangered in every part of its range.



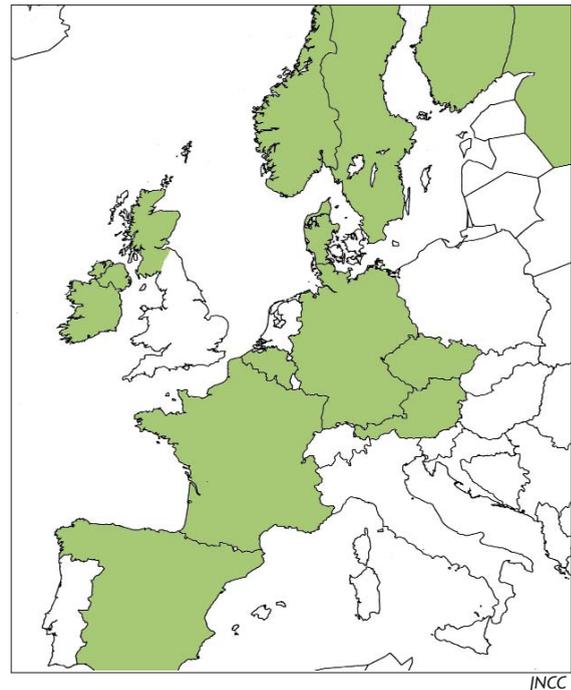
Sue Scott

**The freshwater pearl mussel once provided an important industry, but its numbers have declined dramatically in recent years due to overfishing.**

## Status and distribution

Protected under Schedule 5 of the Wildlife and Countryside Act (1981) and the Wildlife (Northern Ireland) Order 1985, the freshwater pearl mussel is also listed on annexes II and V of the EU Habitats and Species Directive and Appendix III of the Bern Convention. It is included on the IUCN Invertebrate Red List, where its status is described as Vulnerable (IUCN 1990). Classified as a priority species by the UK Biodiversity Steering Group, a national Species Action Plan has been prepared to encourage measures for its survival.

The freshwater pearl mussel is distributed from the Arctic and temperate regions of western Russia through Europe to the northeastern seaboard of North America. Recent studies have revealed that there have been dramatic declines throughout its Holarctic range: for example, Bauer estimates a 95–100% decline in known populations in central and southern Europe (Bauer 1986, 1988). Substantial populations with active recruitment are now found on fewer than 50 rivers in Canada, northwest Russia and northeast Scandinavia, with a handful of sites in Bavaria, the Czech Republic and Austria (Young *et al.* 2000).



JNCC

**The freshwater pearl mussel is found throughout northwest Europe, but in declining numbers.**

## UK distribution

Formerly widespread and abundant in areas of England and Wales, recent surveys revealed most former populations to be virtually extinct, with very little active recruitment (Chesney & Oliver 1998). There have been major declines in Northern Ireland, with recent survey work again showing population declines and scant recruitment (Kerney 1975, Beasley and Roberts 1996), including the rare hard-water



JNCC

**In Britain, the freshwater pearl mussel is only found in viable populations in some Scottish rivers. Other populations do not have active recruitment and could soon become extinct.**

form, *M. margaritifera durrovensis* (Phillips) which is unique to Ireland (Moorkens & Costello 1994).

Originally widely distributed throughout Scotland, a comprehensive survey from 1996 to 1999 revealed that the freshwater pearl mussel is now extinct in most of the lowlands, and scarce everywhere except for a handful of Highland rivers (Cosgrove *et al.* 2000b). If the present rates of extinction continue, it has been estimated that surviving Scottish populations may only persist for a further 25 years. Public disclosure of precise locations is therefore considered unwise due to continued illegal pearl fishing (Cosgrove & Young 1998).

The early causes of decline were undoubtedly pearl fishing (Young *et al.* 2000) and industrial pollution. The current lack of recruitment may be due to a number of factors, including increasing siltation and eutrophication of rivers (Bauer 1983, 1988). The recent declines in migratory salmonids upon which the larvae depend are also giving cause for concern. River engineering for hydro-schemes, flood defence purposes and fishery improvements continues to pose a serious localised threat. Forestry operations, acidification, effluent from fish farms and chemical sheep dip are further threats to the declining populations (Young *et al.* 2000).

Up to half the world's known remaining populations with active recruitment now occur in Scotland (Young *et al.* 2000). Actions taken within the UK will therefore have a direct consequence for the global survival of this species.

## Life history

The freshwater pearl mussel is one of the longest-lived invertebrates known, and individuals can survive for over 100 years (Bauer 1992). The mussels live buried or partly buried in coarse sand and fine gravel in clean, oligotrophic, fast-flowing and unpolluted rivers and streams. They inhale water through their exposed siphons to filter out minute organic particles on which they feed. Where the species was formerly abundant, it is possible that this filtration acted to clarify river water to the benefit of other species, including juvenile Atlantic salmon (*Salmo salar*) and brown or sea trout (*Salmo trutta*) (Zuiganov *et al.* 1994).

The freshwater pearl mussel is typically dioecious, in common with other freshwater bivalves. It matures at 10–15 years, when the length generally exceeds 65 mm. In early summer (June to July), the males shed sperm into the water, and it is inhaled by the females. The fertilised eggs develop in a pouch on the gills for several weeks, and are released from July to September as tiny larvae, measuring 60–70: m (0.6–0.7 mm), known as glochidia. These resemble tiny mussels, but their shells are held apart until they encounter a suitable host, when they snap shut on to the host's gill filaments (Young & Williams 1984).

Each female ejects between 1 and 4 million glochidia in a sudden, highly synchronised event, usually over one to two days (Hastie 2001b). It is likely that a threshold temperature, or other environmental cue, may trigger glochidial release. The proportion of adults producing glochidia is relatively high, varying from 30–60%, even in sparse populations (Young & Williams 1983). Almost all the glochidia are swept away and die, but a few are inhaled by juvenile Atlantic salmon and brown or sea trout. Infective glochidia can remain viable for up to six days (Zuiganov *et al.* 1994), but most attachments probably occur within a few hours (Young & Williams 1984).

Glochidia attached to the gills of juvenile fish encyst, live and grow in the hyper-oxygenated environment until the following spring. They drop off in May and early June, and must land in clean,



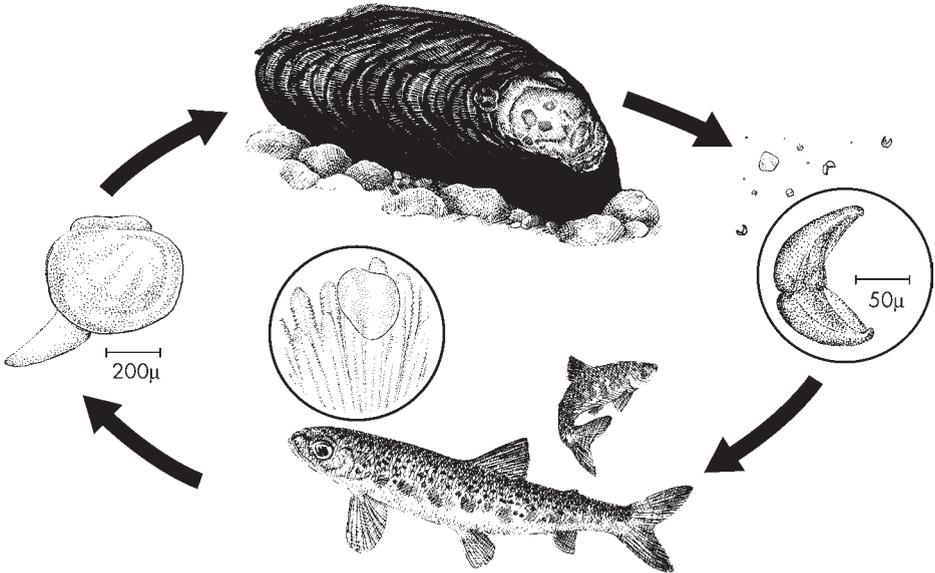
Mark Young

Unattached glochidia.



Mark Young

Encysted glochidia on host gills.



Sarah Wroot

Mussel larvae, known as glochidia, are released in summer and attach themselves to the gill filaments of a juvenile salmonid. Here they encyst and grow until the following spring, when they drop off on to clean gravel and begin maturing. This association does not seem to harm the host fish, and facilitates mussel dispersal.



Mark Young

Newly settled juvenile mussel.



Birgit Heninger

Adult and juvenile mussels from a stable population in Scotland.

sandy or gravelly substrates to settle and start to grow. This association does not appear to harm the fish, and enables young mussels to colonise new areas upstream. Young mussels are typically yellowish-brown, becoming darker with maturity.

Freshwater pearl mussels develop very slowly and can live for more than 100 years, reaching 12–15 cms in length. The life span and maximum size reached are highly variable between populations, depending on environmental conditions, particularly hydrochemistry and water temperature (Zuiganov *et al.* 1994). Freshwater pearl mussel populations tend to be faster growing and shorter lived in the southern part of their range, with a lower reproductive output (Hastie 2001b) than those in the upper (cooler) reaches of catchments and more northerly latitudes (Bauer 1992, Hruska 1992). The mussels can re-bury themselves if dislodged, and can also move slowly across sandy sediments. Most have about a third of their shells exposed, but some adults and virtually all juveniles burrow completely into the substrate and under loose stones (Hastie *et al.* 2000a).

The huge losses involved in this unusual life cycle make the freshwater pearl mussel particularly vulnerable to adverse conditions.

## Population dynamics

Low adult mussel mortality and great longevity historically compensated for the characteristic life cycle of very poor glochidial success and low juvenile recruitment. There is no current scientific agreement about what constitutes a functional mussel population in terms of a viable size/age profile or proportion of juveniles.



Sue Scott/SNH

***M. margaritifera* develops slowly and can live for 100 years. Low adult mortality and longevity compensate for poor glochidial success and low juvenile recruitment.**

Theoretically, age structures in a viable population should reflect both active recruitment and longevity. Adequate recruitment may be indicated by 20% of the population being less than 20 years old (Young *et al.* 2001, Hastie *et al.* 2001b). Very few rivers now show this level of recruitment (Cosgrove *et al.* 2000a, Young *et al.* 2000). Adult mortality of 10% per decade has been suggested as sustainable if recruitment is normal (Bauer 1983, 1986 and 1992). There remains some debate as to the natural rate of recruitment and adult mortality in established populations, and this area requires further research. According to most researchers, sex ratios are typically 1:1 (Bauer 1979). Fertility (the total number of glochidia produced during a lifetime) increases with longevity, so populations with older age classes should be more viable unless juvenile mortality is high. Reduction in aged mussel numbers can be a result of pearl fishing, but higher temperatures, eutrophication and increases in nitrate levels may also reduce longevity. This effect is

exacerbated at low latitudes by naturally reduced life spans (Bauer 1986,1992). The age maximum is therefore different for each population and the target should be as high as is naturally possible.

In Britain, age maxima of less than 40 years impose severe stress on successful recruitment and indicate possible extinction (Hastie *et al.* 2000b). In some rivers, longevity may also be reduced where surrounding limestone strata increases calcium input (Bauer 1992). However, this is a natural phenomenon and should not require remedial action.

Genetic diversity is regarded as low, and most researchers regard the eastern North American populations as the same species as those in Europe. However, virtually nothing is known about the genetic basis of observed population differences in the Margaritiferidae, and this area needs urgent investigation (Chesney & Oliver 1998).

Transplantation of mussels from river to river is relatively unsuccessful, with mortalities of over 50% in the first three years, compared to short-term survival rates of 90% from a translocation exercise within the same river system (Valovirta 1998). There may be either physiological accommodation or a genetic adaptation to particular rivers. The implication is that species recovery plans dependent on introducing new stock may not be successful in the long term. Translocation exercises should therefore be limited to those situations where the threat is particularly severe, and action should be concentrated on maintaining and enhancing all current populations. The longevity of the species means that, given the correct conditions for recruitment, small populations can be sustainable over the long term. **Life in UK Rivers** and Aberdeen University are currently investigating techniques for captive breeding of the species as a means of reinforcing recruitment within catchments.

Current research on population genetics using DNA analyses is underway, and an initial study using RAPD techniques has indicated differences in variability within populations. The results suggest that there are differing factors acting on genetic variation – for example, mussels from larger rivers showed greater variation than those from small streams. The study also showed that Irish populations were more variable than those from the Lake District. This work is continuing (Oliver, pers. comm.).

## Glochidium and host attributes

Only sea trout, brown trout and Atlantic salmon are known to host complete metamorphosis in Europe, where they are the only native host species (Young & Williams 1984). Brown trout are the main host species in Ireland (Beasley 1996) and in Germany, where salmon are now thought to be extinct. American brook charr (*Salvelinus fontinalis*) acts as a host in North America and has been introduced into Europe. However, the rainbow trout (*Oncorhynchus mykiss*) is completely unsuitable (Young & Williams 1984).

*Salmo trutta* exists in two forms: the resident brown trout and the migratory sea trout. Rivers carry varying population ratios, but their relative importance for freshwater pearl mussel has not been studied. The differing reproductive behaviour of these forms most probably affects mussel recruitment. The conservation of both brown and sea trout involves different parameters, and it is therefore necessary to determine the relative importance of the hosts in any one river through appropriate research. 0+ salmon are often more abundant than 0+ trout, and are therefore likely to be the most important hosts of the



Sue Scott

**Atlantic salmon and brown trout are the main hosts for pearl mussel glochidia. Therefore, measures to conserve the mussel must also include the host fish.**

freshwater pearl mussel in most rivers. However, a number of mussel populations in small streams in Scotland have no (or very few) salmon, and these must be considered to be largely trout-dependent.

Host fish become progressively resistant to glochidial infection (Bauer & Vogel 1987, Zuiganov *et al.* 1994). Those in the first three year-classes (but mostly 0+ and 1+ years) form the majority of the host population (Young & Williams, 1984). Older fish may be less susceptible to infection for a number of reasons, including reduced exposure (older fish prefer deeper water away from mussel beds), and acquired immunity due to previous infections (Bauer & Vogel 1987, Zuiganov *et al.* 1994).

The minimum density of fish required to maintain mussel population densities in the long term can theoretically be calculated from a variety of factors, including the fecundity of female mussels; the probability of a glochidium finding a host, of a glochidium reaching metamorphosis, and of young mussels reaching sexual maturity; and the number of generations.

However, the huge mortalities make any such calculations (which are based mainly on laboratory studies and not field data) extremely difficult, and results need to be treated with caution. The figure of 0.2 fish m<sup>-2</sup> given by Bauer (1991) is based on observations of host density and presence of viable mussel populations, and was not calculated. The figure for salmon of 0.05 fish m<sup>-2</sup> from Zuiganov *et al.* (1994) is calculated, but a real figure from the Varzuga River is 0.29 fish m<sup>-2</sup>. An acceptable level of brown trout has been suggested to be in the order of 0.3 fish m<sup>-2</sup> (Zuiganov 1994), but this has not been adequately researched.

Increased glochidial success can theoretically be achieved by increasing host numbers or through the artificial infection of host fish. This strategy should result in more post-glochidial juveniles being introduced into the community, providing mussel numbers are limited by low fish populations rather than lack of suitable habitat. Such measures will only be effective in rivers with appropriate water quality and substrate conditions, such as those where populations have been depleted by over-fishing (Young 1991). Even mild eutrophication can be detrimental to successful reproduction and therefore hamper recovery programmes.

## Habitat requirements

The freshwater pearl mussel life cycle involves an adult stage, living as a filter feeder, a juvenile stage living interstitially in sediment, and a larval (glochidial) stage living attached to the gills of trout or salmon. All life stages therefore need consideration, as does the viability of the host species of fish. Adults are more tolerant of a wider range of in-river conditions than juveniles (Hastie *et al.* 2000a).

## Water quality

Water chemistry has received much attention and is widely quoted in the literature when environmental parameters for optimum mussel survival are under discussion. Bauer (1988) is most often quoted, and there is a general acceptance that the pearl mussel prefers oligotrophic conditions – poor in nutrients, pH 7.5 or less, and with low overall conductivity. The actual figures cited by Bauer were based on the correlation of field conditions to mussel presence or absence in central European rivers.

Similar correlations have been noted in the UK but exceptions do exist. The target levels given here reflect the current situation in the Scottish rivers where the populations are large and recruitment is present. The situation in English rivers with large adult populations and scant recruitment is currently unfavourable, and no conclusions can therefore be drawn from the present water quality. A few atypical populations in England and Ireland appear to be adapted to tolerate more calcareous water chemistry, where the surrounding geology increases calcium content beyond the levels suggested by Bauer.

Pearl mussels are most vulnerable to human influences at the stage where they leave the host fish and establish in the sediment. This stage will die out completely if even a slight degree of pollution is present. The juveniles are also far less tolerant than the adults, and persistent intermediate levels of

eutrophication could prevent long-term recruitment, resulting in aged stocks (Bauer 1988).

The critical parameters affecting recruitment are BOD (biochemical oxygen demand), calcium and phosphate levels in the water. Bauer (1988) observed that adult mortality was correlated with nitrate concentration, and that increased levels of phosphate, calcium and BOD were correlated with decreasing survival and establishment of juveniles.

Research has indicated that nitrate levels must not exceed  $1.0 \text{ mg l}^{-1}$ , although higher values of  $1.5 \text{ mg l}^{-1}$  may be encountered in some British rivers.

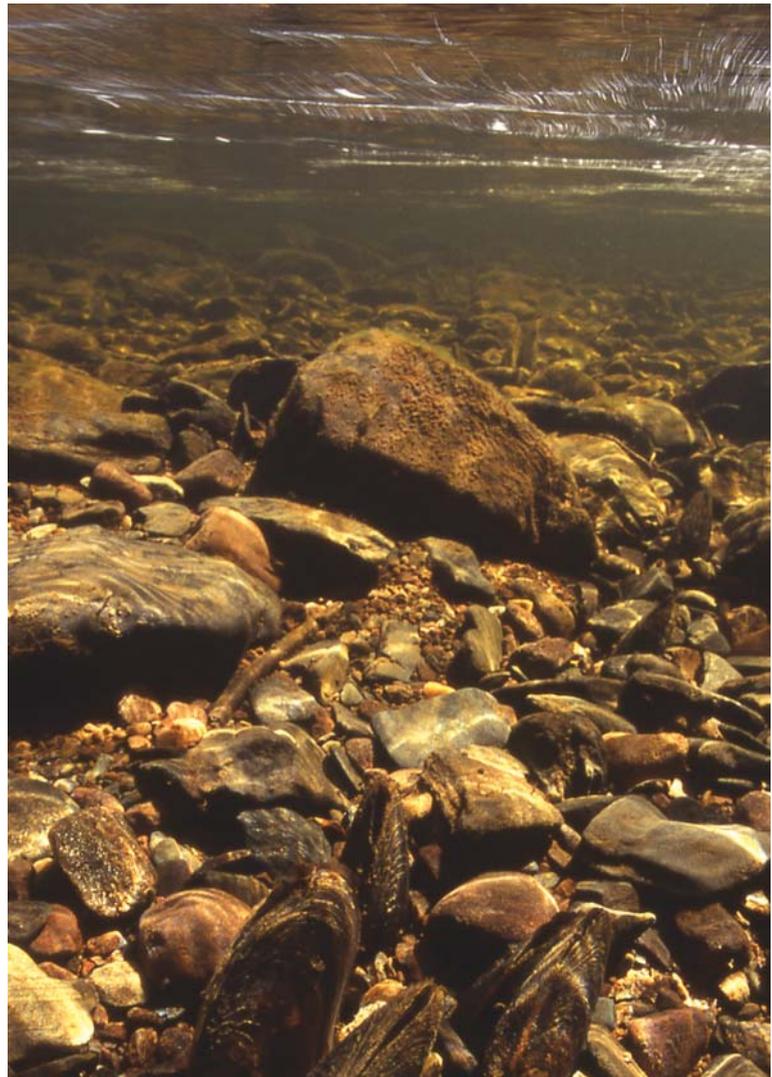
Phosphates should be  $<0.03 \text{ mg l}^{-1}$ . Conductivity must not exceed  $100 \mu\text{S cm}^{-1}$ , although higher values of  $120 \mu\text{S cm}^{-1}$  may be natural on limestone-influenced stretches (Oliver 2000).

BOD and dissolved oxygen are therefore undoubtedly of importance to pearl mussel survival. These parameters are currently measured by sampling the water column, and are therefore relevant only to the adult phase. Juvenile mussels live interstitially, but no monitoring of substrate oxygen levels is currently undertaken. The levels tolerated by juveniles have only been measured by Buddensiek *et al.* (1993) for four rivers in Germany. Monitoring and research on the interstitial environment are therefore urgent priorities.

## Water quantity

The influence of stream hydrological processes on microhabitat, particularly hydrodynamic effects on juvenile recruitment, is poorly understood. Most researchers report minimum/maximum depths and velocities for *M. margaritifera*, *M. laevis* and *M. falcata* within the ranges of 0.1–2 m and  $0.1\text{--}2 \text{ m s}^{-1}$ . (Vannore & Minshall, 1982). Optimum depth and velocity in Scottish rivers were found by Hastie *et al.* (2000a) to be 0.3–0.4 m deep and  $0.25\text{--}0.75 \text{ m s}^{-1}$ . Work on complex characteristics of flow has shown shear stress over existing mussel beds to be a major factor in determining recruitment in some mussel species in the USA, but this work did not include the freshwater pearl mussel (Layzer & Madison 1995).

No absolute figures are available for a minimum suitable flow velocity. Until such factors have been quantified by further research, a precautionary approach should be followed. Low summer flow velocities can allow the formation of algal mats and reduce interstitial–water column mixing. The



Sue Scott

**The water over a pearl mussel bed must be clear and well-oxygenated. Maximum depth is about 2 m.**

uncovering of shallow riffle areas and the aggregation of detrital silt are indicators of poor conditions for both adult and juvenile mussels. Water flow in summer should therefore be sufficient so as not to induce low oxygen levels or heat stress, and sufficient to reduce the sedimentation of fine particles and detritus, especially in areas where juveniles aggregate. Research into the effects of flow velocities (including summer spates) on pearl mussel habitat is urgently required.

Moderate flooding may have a positive effect in cleaning silts from gravel beds and riffles. Autumn flows can wash out algal mats and sediments accumulated over the summer, but severe floods can remove large numbers of mussels from their beds and are potentially disastrous for populations where recruitment is not taking place (Hastie *et al.* 2001b). This study showed that an estimated 1:100-year return period flood directly killed between 4–8% of the population due to significant channel reformation and large-scale movements of substrata. Climate change appears to be resulting in increased frequency of high flow occurrence and greater annual run-off events (Black 1996), which may be a cause for future concern.

Even slight hydrological changes may result in serious degradation of habitat due to the very specific sediment requirements of juveniles (Bauer 1988, Buddensiek *et al.* 1993). Catchments should therefore be monitored for alterations to land drainage, and all work that has the potential to increase siltation or the speed of run-off should be subject to rigorous examination and environmental impact assessment. Regulated rivers with alternating high and low flows (or those where regulation is proposed) should give particular cause for concern if they support thriving populations with active recruitment.

## Substrate

The characteristics of riverbed substrata are of critical importance for freshwater pearl mussel populations. The typical substrate preference is small sand patches stabilised amongst large stones or boulders in fast-flowing streams and rivers. Such boulder-sheltered mussel beds may be critical for



Sue Scott

**Typical substrate preference of the freshwater pearl mussel is large boulders and cobbles, with patches of sand in-between.**

(Hastie 2001b). Juveniles are thought to require sediment of low organic content for their further development (Bauer *et al.* 1980), with a structure that allows a high rate of exchange between the free water body and the interstitial water. Oxygen, pH and ammonia appear to be important chemical parameters in this respect (Buddensiek *et al.* 1993).

Gradient could affect mussel distribution indirectly by determining the stability of the substrata. Purser (1985) demonstrated that an intermediate gradient range of 0.8–3 m km<sup>-1</sup> was preferred. The majority of adult mussels live in dense beds in substrates of mixed cobble, stone and sand at the tail-end of pools or in the moderate flow channels of river bends. In these sites, stability of the bed is important.

recruitment after heavy floods (Vannote & Minshall 1982). Riffle areas with mixtures of rocks, cobbles and sand are important habitats in low-gradient sections, providing a well-oxygenated and silt-free environment. Juveniles are mostly associated with such riffles, and require fine sediment within which to shelter.

There is consensus among researchers that the early post-settlement period, when juveniles establish themselves in sediment, is the most sensitive and critical phase in the pearl mussel life cycle

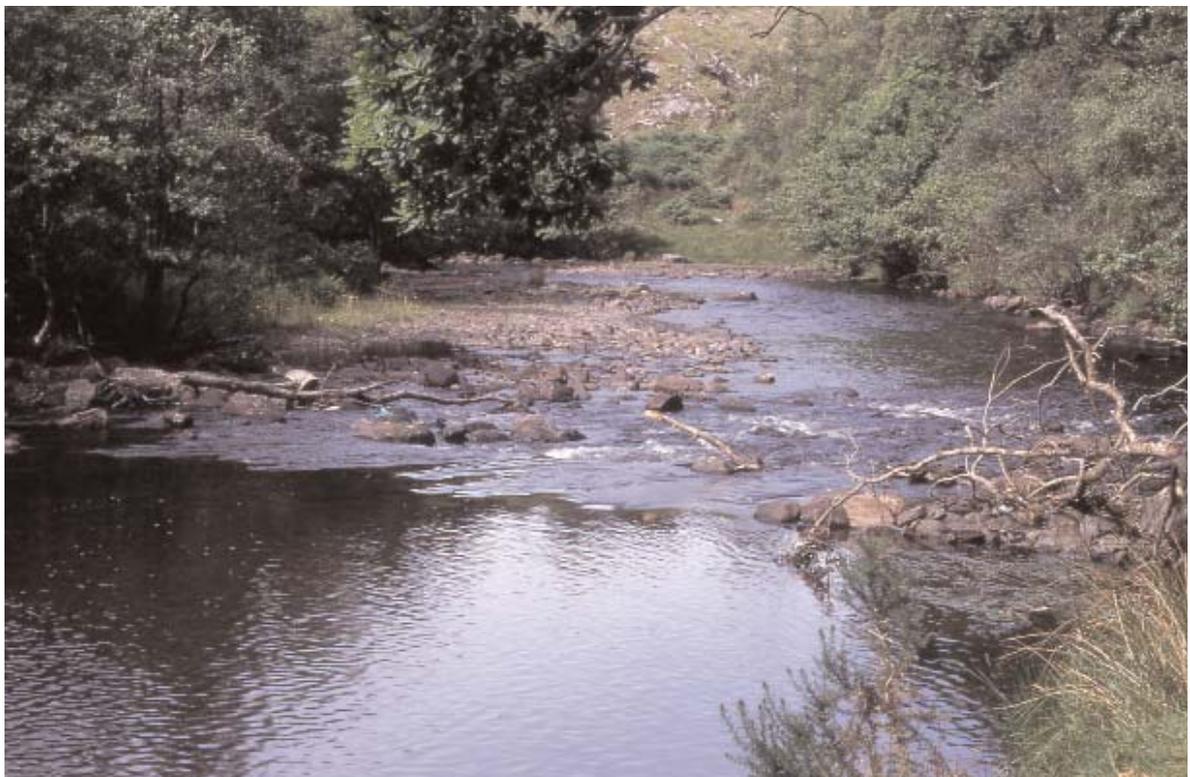
By contrast, loose sands and gravels on the inner curves of river channels are rarely inhabited by mussels because of their instability (Hastie *et al.* 2000a). Densely vegetated areas are unlikely to be suitable, since these tend to trap silt and organic debris.

Siltation of suitable substrates is therefore a severe problem that can be caused by increased sediment load and detrital production due to eutrophication. While bank erosion, flooding and land drainage can all affect sediment load and threaten adult mussel beds, even small amounts of sediment can alter the interstitial environment of the juveniles. If the interstitial spaces are clogged, the young mussels suffocate (Hastie *et al.* 2000a). The level of  $30 \text{ mg l}^{-1}$  of suspended solids has been noted by Valovirta (1998) as the limit of tolerance by adult mussels. This level may not be critical if it occurs for a short time during floods. However, long-term levels of suspended solids should be much less. Levels consistently above  $10 \text{ mg l}^{-1}$  should give cause for concern.

Stable gravel/cobble and riffle sites should therefore have very low levels of silt in the interstitial spaces of the substrate, but this has not been quantified by field research. Research is urgently needed into the effects of and tolerance to interstitial siltation.

### Channel structure and management

Mussel aggregations in many of the rivers surveyed in England and Wales are associated with areas of shade, normally created by overhanging herbaceous vegetation, scrub and bank-side trees, with little or no bank erosion. Shade keeps water temperatures down during the summer months and inhibits the growth of filamentous algae. Algal mats overgrowing mussel beds can impair respiration, feeding, fertilisation and the release of glochidia (Hastie 2001b). In Scotland many populations occur in areas not currently forested. It is therefore suggested that tree cover in itself is not necessary where nutrient levels are low enough to prohibit algal blooms, and flows are sufficient to prevent high summer temperatures. Stable channels with little bed transport except in floods are important features, and channel structure should not be altered in any way that will impede water flow, increase flooding, or alter the distribution of substrates.



Sue Scott

**Mussel rivers ideally have areas of riffle, with overhanging vegetation providing shade.**



Sue Scott

**River engineering schemes that reduce water levels can be lethal for pearl mussels, as can siltation and nutrient enrichment.**

such as dredging, canalisation, scouring and weir construction cause alteration or loss of suitable river bed substrata (Young & Williams 1983). Any such activities or proposals should therefore be subject to rigorous environmental assessment.

## Other factors

### Host fish stocks

The long-term survival of the freshwater pearl mussel depends ultimately upon host availability. There is concern that significant changes in salmon and sea trout stocks may threaten populations (Cosgrove *et al.* 2000a, Chesney & Oliver, 1998). Salmon catches in northwest Scotland, where the majority of the pearl mussel populations are concentrated, exhibited a marked decline during the 1990s and are now at historically low levels. Several migratory trout stocks have collapsed, and sea trout catches are now at the lowest levels ever recorded (Hastie & Cosgrove 2001). The cause has been attributed to many factors, including climate change, over-fishing and predation, infestation by sea lice (now unequivocally linked to the decline in sea trout stocks), acidification and degradation of habitat (Marshall 1998).

It is widely accepted that eutrophication is a major cause of mussel decline and that agricultural practices, particularly efficient field drainage and the use of fertilisers, are major contributors to this process. To reduce this effect, a buffer of natural vegetation may promote the removal of nutrients. Costello *et al.* (1998) suggested a buffer zone of 10 m with intercepted field drains, but effective size will depend on soil type and slope (and therefore vulnerability to erosion), river width and catchment land use. Cattle drinks and poached banks are undesirable features as they cause additional siltation.

The importance of surrounding vegetation has been stressed by Hruska (1996), not only in ameliorating nutrient run-off but also as a supplier of food for the juvenile stage. Hruska suggests that surrounding areas of wet grass meadows are most beneficial to juvenile growth. Whilst of relevance to some rivers in the UK, the situation is different in Scotland, and this study needs careful consideration if its findings are to be applied in the UK.

River engineering for hydro-electric schemes, flood protection or fisheries reasons may cause local extinctions (Young 1995). Activities

It is not known whether brown trout produce enough fry to sustain Scottish mussel populations over the long term, and there is an urgent need to know more about the relationship between host stock sizes and the reproductive success of mussels (Chesney & Oliver 1998). It is possible that there is a mutually beneficial relationship between the freshwater pearl mussel and host fish. Adult mussels can filter up to 50 litres of water per day (Zuiganov *et al.* 1994), and mussel beds may provide a microhabitat for invertebrates on which juvenile salmon feed (L Hastie pers.obs.).

Research is urgently needed to investigate the coincidental decline in salmonid fish stocks and the freshwater pearl mussel. **Life in UK Rivers** and Aberdeen University are currently undertaking work that will enable models to be developed relating mussel recruitment to the age and species of salmonid fish. This should assist in the production of guidance for appropriate fish management. It is also important to ensure that there are no barriers to fish migration and no acidification of tributaries, as these factors could have an adverse effect on salmonid fry availability.

Disturbance while electrofishing to assess salmonid stocks appears to have no short-term adverse effect on pearl mussel populations (Hastie & Boon 2001). However, regular trampling should be kept to a minimum.

## Eutrophication

Filamentous algal mats, high phytoplankton production and the detritus formed by their decay indicate undesirable levels of eutrophication, with the resultant inhibition of both juvenile success and adult well being. A build-up in summer can also indicate inadequate flow rates. The appearance of obvious algal production is therefore an indicator of poor river condition and should warn of environmental conditions that are deleterious to freshwater pearl mussel populations.

## Inorganic pollutants

Pyrethroid sheep dip is known to be highly toxic to freshwater arthropods, but the direct effect on freshwater pearl mussels is not known. Indirect effects could occur through a reduction in salmonid fish numbers.

## Acidification

Acidification is known to have deleterious effects on juvenile trout and salmon, and could therefore have an indirect effect on freshwater pearl mussels. Acidification may also alter juvenile habitat. According to Meyer & Klupp (1980) the decline in pearl mussels in some German rivers may be due to lowered pH values, exacerbated by spruce monocultures. The practice of catchment liming as a counter measure to restore salmonid habitat may also have an adverse effect, given the sensitivity of pearl mussels to high calcium levels.

## Introduced species

Mussel viability is dependent on host salmonid viability, so any threats to native trout and salmon stocks should be avoided. There has been a tendency for angling societies and individuals to release non-native salmonids such as rainbow and brook trout. These species are a potential threat to the long-term survival of the freshwater pearl mussel as they may out-compete the native species and do not act as glochidial hosts.

## Priorities for research

- The effects of flow rate on habitat, particularly substrate.
- The effects of eutrophication (and water quality requirements, especially in England).
- The effects of siltation.
- Genetic variation.

- The importance of different host species and appropriate densities.
- Key aspects of the interstitial environment for juveniles.
- Techniques for artificially rearing juveniles.
- Tolerance to acidity and sheep dip.
- Recruitment processes and rates, related to growth and water temperature (and therefore climate change).

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## References

- Bauer G (1979). Untersuchungen zur Fortpflanzungsbiologie der Flussperlmuschel im Fichtelgebirge. *Archiv für Hydrobiologie* 85, 152–165.
- Bauer G, Schrimppf E, Thomas W and Herrmann R (1980). Zusammenhänge zwischen dem Bestanddruckgang der flussperlmuschel (*Margaritifera margaritifera*) in Fichtelgebirge und der Gewässerbelastung. *Archiv für Hydrobiologie* 88, 505–513.
- Bauer G (1983). Age structure, age-specific mortality and population trend of the freshwater pearl mussel in N. Bavaria. *Archiv für Hydrobiologie* 98, 523–532.
- Bauer G (1986). The status of the freshwater pearl mussel *Margaritifera margaritifera* L. in the south of its European range. *Biological Conservation* 38, 1–9.
- Bauer G & Vogel C (1987). The parasitic stage of the freshwater pearl mussel *Margaritifera margaritifera* L. Host response to glochidiosis. *Archiv für Hydrobiologie* 76, 393–402.
- Bauer G (1988). Threats to the freshwater pearl mussel in Central Europe. *Biological Conservation* 45, 239–253.
- Bauer G (1991). Plasticity in life history traits of the freshwater pearl mussel. In: Seitz A & Loeschcke W (eds). *Species Conservation: A population biological approach*. Birkhauser Verlag, Basel. 103–120.
- Bauer G (1992). Variation in the life span and size of the freshwater pearl mussel. *Journal of Animal Ecology* 61, 425–436.
- Beasley CR (1996). *The distribution and ecology of the freshwater pearl mussel Margaritifera margaritifera L. 1758 in County Donegal, Ireland, and implications for its conservation*. Unpublished PhD Thesis, Queen's University, Belfast.
- Beasley CR and Roberts D (1996). The current distribution and status of the freshwater pearl mussel *Margaritifera margaritifera* L. in northwest Ireland. *Aquatic Conservation: Marine and Freshwater Ecosystems* 6, 169–177.
- Black AR (1996). Major flooding and increased flood frequency in Scotland since 1988. *Physical and Chemical Earth* 5, 463–468.
- Buddensiek V (1995). The culture of juvenile freshwater pearl mussels in cages: A contribution to conservation programmes and the knowledge of habitat requirements. *Biological Conservation* 74, 33–40.
- Buddensiek V, Engel H, Fleischauer-Rossing S, and Wachtler K (1993). Studies on the chemistry of interstitial water taken from defined horizons in the fine sediments of bivalve habitats in several North German lowland waters II: microhabitats of *Margaritifera margaritifera* L. *Archiv für Hydrobiologie*, 127, 151–166.
- Chesney HCG & Oliver PG (1998). Conservation issues for Margaritiferidae in the British Isles and Western Europe. In: Killeen I and Holmes AM (eds). *Molluscan Conservation. A strategy for the 21st Century*. Journal of Conchology Special Publication 2, 231–242.
- Cosgrove PJ and Young M (1998). *The status of the freshwater pearl mussel Margaritifera margaritifera in Scotland*. A report to Scottish Natural Heritage, Edinburgh.
- Cosgrove PJ, Hastie LC and Young MR (2000a). Freshwater pearl mussels in peril. *British Wildlife* 11, 340–347.
- Cosgrove PJ, Young MR, Hastie LC, Gaywood M and Boon PJ (2000b). The status of the freshwater pearl mussel *Margaritifera margaritifera* L. in Scotland. *Aquatic Conservation: Marine and Freshwater Ecosystems* 10, 3, 197–208.
- Costello M, Moorkens E, Larkin E, Kurz M & Dowse J (1998). Management priorities for the River Nore (Ireland) to conserve the pearl mussel *Margaritifera margaritifera durrovensis*. *Journal of Conchology Special Publication* 2, 257–264.
- Hastie LC, Boon PJ, Young MR and Way S (2001). The effects of a major flood on an endangered freshwater mussel population. *Biological Conservation* 98, 107–115.
- Hastie LC, Young MR, Boon PJ, Cosgrove PJ and Henninger B (2000b). Sizes, densities and age structures of Scottish *Margaritifera margaritifera* populations. *Aquatic conservation: Marine and Freshwater Ecosystems* 229–247.
- Hastie LC, Boon PJ & Young MR (2000a). Physical microhabitat requirements of freshwater pearl mussel *Margaritifera margaritifera* (L.). *Hydrobiologia* 429 59–71.
-

- Hastie LC and Boon PJ (2001a). Does electrofishing harm freshwater pearl mussels? *Aquatic conservation: Marine and freshwater Ecosystems* 11, 149-152.
- Hastie LC and Cosgrove PJ (2001). The decline of migratory salmonid stocks: a new threat to pearl mussels in Scotland. *Freshwater Forum* 15, 85–96.
- Hastie LC, Young MR and Boon PJ (2000). Growth characteristics of freshwater pearl mussels *Margaritifera margaritifera* (L.). *Freshwater Biology* 43, 243–256.
- Hastie LC and Young MR (2001). Freshwater pearl mussel *Margaritifera margaritifera* glochidiosis in wild and farmed salmonid stocks in Scotland. *Hydrobiologia* 445, 109–119.
- Hruska J (1996). The freshwater pearl mussel in South Bohemia: evaluation of the effect of temperature on reproduction, growth and age structure of the population. *Archiv fur Hydrobiologie* 126, 181–191.
- Kerney MP (1975). European distribution maps of *Pomatus elegans*, *Discus ruderatus*, *Eobania vermiculata* and *Margaritifera margaritifera*. *Archiv fur Molluskenkunde* 106, 243–249.
- Layzer JB and Madison LM (1995). Microhabitat use by freshwater pearl mussels and recommendations for determining their instream flow needs. *Regulated Rivers: Research and Management* 10, 329–345.
- Marshall S (1998). West Sutherland Fisheries Trust (WSFT) Annual Review 1997. West Sutherland Fisheries Trust, Inverness. 19pp
- Meyer L and Klupp R (1980). Ueber die situation der perlmuschel in Oberfranken. *Fischerei und Teichwirt* 317, 194–197.
- Moorkens EA and Costello MJ (1994). Imminent extinction of the Nore freshwater pearl mussel *Margaritifera durrovensis* Phillips: a species unique to Ireland. *Aquatic Conservation: Marine and Freshwater Ecosystems* 4, 363–365.
- Oliver PG (2000). *Conservation Objectives for the freshwater pearl mussel Margaritifera margaritifera L.* Report for English Nature, Peterborough.
- Purser GJ (1985). *Factors affecting the distribution of the freshwater pearl mussel Margaritifera margaritifera L. in Britain.* Unpublished PhD Thesis, University of Aberdeen, Aberdeen.
- Valovirta I (1998). Conservation of *Margaritifera margaritifera* in Finland. Coll. On the Bern Convention. *Environmental Encounters* 109, 59–63.
- Vannote RL and Minshall GW (1982). Fluvial processes and local lithology controlling abundance, structure and composition of mussel beds. *Proceedings of the National Academy of Science* 79, 4103–4107.
- Young MR (1991). Conserving the freshwater pearl mussel *Margaritifera margaritifera* L. in the British Isles and continental Europe. *Aquatic Conservation: Marine and Freshwater Ecosystems* 1, 73–77.
- Young MR and Williams J (1983). The status and conservation of the freshwater pearl mussel in Great Britain. *Biological Conservation* 25, 35-52.
- Young MR and Williams J (1984). The reproductive biology of the freshwater pearl mussel in Scotland I & II. *Archiv Fur Hydrobiologie* 99, 405–422, and 100, 29–42.
- Young MR (1995). *The distribution, ecology and conservation of the freshwater pearl mussel (Margaritifera margaritifera L.) in Scotland.* Information and Advisory Note No. 2, Scottish Natural Heritage, Edinburgh.
- Young MR, Cosgrove PJ and Hastie LC (2000). The extent of, and causes for, the decline of a highly threatened naiad: *Margaritifera margaritifera*. In: Bauer G and Wachtler K (eds). *Ecological Studies Vol 145. Ecology and Evolutionary Biology of the freshwater mussels Unionoidea.* Springer-Verlag, Berlin.
- Young MR, Hastie LC and al-Mousawi B (in press). What represents an “ideal” population profile for *Margaritifera margaritifera*? In: Bauer G (ed.) *The Freshwater Pearl Mussel in Europe.* Wasserwirtschaftsamt, Hof, Germany.
- Zuiganov V, Zotin A., Nezlin L & Tretiakov V (1994). *The freshwater pearl mussels and their relationships with salmonid fish.* VNIRO, Moscow. 104 pp.
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Life in UK Rivers was established to develop methods for conserving the wildlife and habitats of rivers within the Natura 2000 network of protected European sites.

Set up by the UK statutory conservation bodies and the European Commission's LIFE Nature programme, the project has sought to identify the ecological requirements of key plants and animals supported by river Special Areas of Conservation.

In addition, monitoring techniques and conservation strategies have been developed as practical tools for assessing and maintaining these internationally important species and habitats.



The freshwater pearl mussel is one of the longest-lived invertebrates, sometimes surviving for over 100 years, and has a complex, fascinating life cycle. It has developed a unique relationship with trout and salmon that involves its larvae attaching to the host fish's gill filaments where they develop.

Affected by reduced water quality and increased siltation, the freshwater pearl mussel is also made vulnerable by its association with trout and salmon to declines in the populations of these species. As a result, its numbers have decreased dramatically across Europe. In the UK, viable populations are found only in a number of Scottish sites that now hold almost half the world's remaining populations with active recruitment.

The freshwater pearl mussel is one of the species at the heart of a major European effort to conserve key freshwater animals and plants and the river habitats that sustain them. This report describes the ecological requirements of this important species in a bid to assist the development of monitoring programmes and conservation strategies that are vital for its future.

Information on Conserving Natura 2000 Rivers and Life in UK Rivers can be found at [www.riverlife.org.uk](http://www.riverlife.org.uk)

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